



PPARgamma ELISA Kit (Colorimetric)

Catalog Number TE-0009

(For Research Use Only)

Introduction

Peroxisome-proliferator-activated receptor gamma (PPAR gamma) is a ligand-activated transcription factor that plays an important role in the control of gene expression associated with a variety of physiological processes, particularly in metabolism and adipogenesis. With ligand binding, PPAR gamma forms a heterodimer with the retinoic X receptor, and binds to PPAR response elements in the promoter region of target genes, thus regulating their expression. Dysfunction of PPAR gamma leads to the pathological processes, such as metabolic diseases and cancer. Furthermore, modulation of receptor action in these diseases may be of therapeutic value. PPARgamma has been demonstrated to be the target of the thiazolidinediones (TZDs), which are widely used to treat type 2 diabetes. Signosis has developed PPARgamma ELISA, to help specific and sensitive measurement of the activation of PPARgamma.

Principle of the assay

PPARgamma ELISA kit is high sensitive and specific assay with a simple and optimized procedure. The 96-well (8X12 strip) clear plate is pre-immobilized with the PPARgamma consensus sequencing oligo. The activated PPARgamma in nuclear extract or the whole cell lysate is added in the well and binds to the oligo. The activated PPARgamma is detected with a specific antibody against PPARgamma subunit and a HRP conjugated secondary antibody. The assay utilizes colorimetric detection method, which can be easily measured by spectrophotometry.

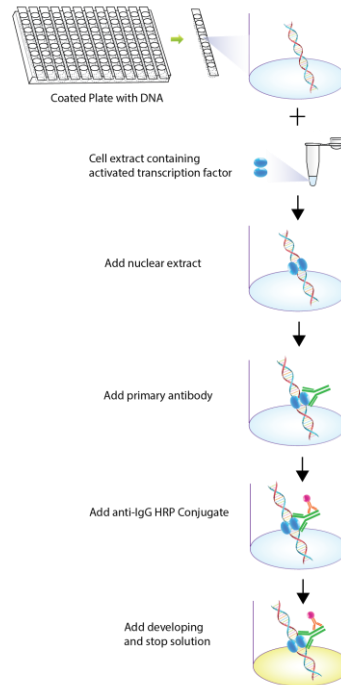


Diagram of TF ELISA

Materials provided with the kit

- 96 well microplate coated with PPARgamma consensus oligo (4°C).
- Antibody against PPARgamma (4°C).
- HRP conjugate secondary antibody (4°C)
- 2X TF binding buffer (-20°C).
- 1X Nuclear extract dilution buffer (-20°C).
- PPARgamma Positive control (-20°C)
- 1X Diluent buffer (4°C)
- 5X Assay wash buffer ((4°C)
- Substrate (4°C)
- Stop Solution (4°C)

Material required but not provided

- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer
40ml 5x Assay wash buffer
160ml ddH₂O
- Dilute 500 times of antibody against PPAR γ with 1X Diluent buffer before use.
- Dilute 1000 times of HRP conjugate secondary antibody with 1X Diluent buffer before use.

Assay procedure

1. Cut the sealing film over the plate and remove it from the desired number of well strips. Make sure the rest of wells are well sealed.
2. Make TF binding mix
25ul 2X TF binding buffer
X Nuclear extract (2-10ug)
X Nuclear extract dilution buffer
Total 50ul
For positive control, use 25ul of positive control without adding nuclear extract dilution buffer.
3. Add the mix on a well and incubate for 1 hour with gently shaking at room temperature.
4. Discard the contents and wash by adding 200 μ l of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Complete removal of liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.
5. Add 100 μ l of diluted antibody against PPAR γ to each well and incubate for 1 hour at room temperature with gentle shaking.
6. Repeat the aspiration/wash as in step 4.
7. Add 100 μ l of diluted HRP conjugate secondary antibody to each well and incubate for 45 min at room temperature with gentle shaking.
8. Repeat the aspiration/wash as in step 4.
9. Add 100 μ l of substrate to each well and incubate for 15-30 minutes.
10. Add 50 μ l of stop solution to each well. The color in the wells should change from blue to yellow.
11. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.